

## **GROUNDWATER MONITORING REPORTING REQUIREMENTS**

MONITORING REPORTS (QUARTERLY, OR SEMI ANNUALLY)

Information submitted to MDA should include:

- 1. Analytical results for samples including QA samples;
- 2. Stabilization, purge volume and/or recovery rate test forms;
- 3. Copies of "chain-of-custody" forms, lab analytical reports, lab QA/QC reports, and MDA Laboratory Data Review Checklist (GD29 Attachment);
- 4. Copies of field records;
- 5. Water level measurements to the nearest 0.01 foot from the surveyed point referenced to the National Geodetic Vertical Datum or an identified benchmark;
- 6. A table showing all water level information to date;
- 7. A table showing all analytical results to date. Include numerical information (specify reporting limit) rather than "non detect" for those parameters not detected above reporting limits;
- 8. A detailed site map showing the location of all monitoring wells with ground water elevation contours. The map should clearly indicate the elevations of ground water contour lines and ground water elevations in each well. The map should also indicate expected direction of ground water flow with a directional arrow. All maps should have a north arrow and scale;
- 9. A brief description of how the ground water contour map was completed (for example, the name of the ground water model used);
- 10. A statement explaining the reasons for and ramifications of any deviation in sampling, analytical techniques or equipment from those stated above or in the laboratory quality assurance/quality control plan:
- 11. A hydrograph showing changes in water level elevations over time;
- 12. Provide all calculations needed to determine unionized ammonia concentrations, if required, and
- Provide all calculations needed to determine the horizontal hydraulic conductivity from slug or plug test data.

### ANNUAL MONITORING REPORT

Once a year, or as agreed to by MDA, a summary and discussion of the monitoring results should be submitted to the MDA. This annual summary should:

- 1. Identify recent and long-term trends in the concentrations of monitored constituents and in water elevations;
- 2. Tabulate the analytical results to date and highlight those that exceeded ground water regulatory standards (such as Health Risk Limits, Health Based Values or Maximum Contaminant Levels). On tables, include numerical information such as <0.1 rather than "Not Detected" for those parameters not detected above reporting limits and list the regulatory standards;
- 3. Provide concentration vs. time graphs for selected analytical parameters. Use an appropriate scale so that all data is easily seen;
- 4. Provide a hydrograph showing changes in water level elevations over time;
- 5. Evaluate the effect the facility is having on ground water;
- 6. Suggest any additions, changes, or maintenance needed in the monitoring program; and
- 7. Provide a ground water flow map (see #8 above).



# CHAIN OF CUSTODY RECORD

| MSD   |                     | PROJE           | PROJECT LEADER   | REMARKS   |             |           |                      |                           |     |
|---|---------------------|-----------------|--|---|-------------|-----------|----------------------|---------------------------|-----|
| PROJECT NAME/LOCATION   | OCATION             |                 |  |   |             |           |                      | lab                       | LAB |
| ESD SAMPLE TYPES 2 GOLA WIRTS 2 GOLA WIRTS 4 WASTEWIRTS 3 MASTER 4 WASTEWIRTS 5 LEJONE 11 OTHER | S SEDIMENT SE       | SAMPLER         | ole R  | DATA TO: CIRCLE/ADD parameters desired list no. of containers submitted | 300         | ANALYSIS  |                      | nbon receipt by           | USE |
| STATION 19  | COMP                | BARD            | STATION LOCATION/DESCRIPTION   | TOTAL CONTAINER:  | AOV ANETALS | BOINDED   | TAG NO!REMARKS       | Custody seals in Seals in |     |
|   |                     |                 |  |   |             |           |                      |                           |     |
|   |                     |                 |  |   |             |           |                      |                           |     |
|   | _                   |                 |  |   |             |           |                      |                           |     |
|   |                     |                 |  |   |             |           |                      |                           |     |
|   |                     |                 |  |   |             |           |                      |                           |     |
|   |                     |                 |  |   |             |           |                      |                           |     |
|   |                     |                 |  |   |             |           |                      |                           |     |
|   |                     |                 |  |   |             |           |                      |                           |     |
|   |                     |                 |  |   |             |           |                      |                           |     |
|   |                     |                 |  |   |             |           |                      |                           |     |
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| (NDIS)  |                     |                 | (SIGN)   | (SIGN)  |             |           | (SIGN)               |                           |     |
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White and Yellow copies accompany sample shipment to Laboratory; Yellow White copy is returned to samplers; Pink copy returned by samplers

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## Containers, Preservation Techniques and Holding Times

| Parameter                               | Container            | Preservative <sup>1</sup>                        | Maximum Holding<br>Time <sup>2</sup>                    |
|---|----------------------|--|---|
| Nitrate + Nitrite                       | 100 ml. poly         | cool 4°C, H <sub>2</sub> SO <sub>4</sub> to pH<2 | 28 days   |
| Ammonia                                 | 100 ml. poly         | cool 4°C, H2SO4 to<br>pH<2                       | record pH and temp.<br>at time of collection<br>28 days |
| Base Neutral Pesticides<br>(MDA List 1) | 1000 ml. amber glass | cool 4°C   | follow applicable method holding times                  |
| Acid Pesticides<br>(MDA List 2)         | 1000 ml. amber glass | cool 4°C, H <sub>2</sub> SO <sub>4</sub> to pH<2 | follow applicable method holding times                  |
| Carbamates<br>(MDA List 3)              | 1000 ml. amber glass | cool 4°C, 5 mls. of 1% phosphoric acid           | follow applicable method holding times                  |
| Pentachlorophenol                       | 1000 ml. amber glass | cool 4°C, H <sub>2</sub> SO <sub>4</sub> to pH<2 | 7 days until extraction                                 |

## **NOTES:**

<sup>&</sup>lt;sup>1</sup>Sample preservation should be performed immediately upon sample collection.

<sup>&</sup>lt;sup>2</sup>Samples should be analyzed as soon as possible after collection. The times listed are the **maximum** holding times that samples may be held before analysis and still be considered valid.



Advantages and Disadvantages of Various Groundwater Sampling Devices (modified from EPA/540/P-91/007 January 1991)

| Device                          | Advantages  | Disadvantages  |
|---------------------------------|---|--|
| Bailer                          | <ul> <li>The only practical limitations are size and materials</li> <li>No power source needed</li> <li>portable</li> <li>Inexpensive; it can be dedicated and hung in a well reducing the chances of crosscontamination</li> <li>Minimal outgassing of volatile organics while sample is in bailer</li> <li>Readily available</li> <li>Removes stagnant water first</li> <li>Rapid, simple method for removing small volumes of purge water</li> </ul> | Time consuming, especially for large wells     Transfer of sample may cause aeration   |
| Submersible Pump                | <ul> <li>Portable; can be used on an unlimited number of wells</li> <li>Relatively high pumping rate (dependent on depth and size of pump)</li> <li>Generally very reliable; does not require priming</li> </ul>  | <ul> <li>Potential for effects on analysis of trace organics</li> <li>Heavy and cumbersome, particularly in deeper wells</li> <li>Expensive</li> <li>Power source needed</li> <li>Susceptible to damage from silt or sediment</li> <li>Impractical in low yielding or shallow wells</li> </ul> |
| Non-Gas Contact<br>Bladder Pump | Maintains integrity of sample     Easy to use   | <ul> <li>Difficult to clean although dedicated tubing and bladder may be used</li> <li>Only useful to approximately 100 feet in depth</li> <li>Supply of gas for operation (bottled gas and/or compressor) is difficult to obtain and is cumbersome</li> </ul>                                 |
| Suction Pump                    | Portable, inexpensive, and readily available  | <ul> <li>Only useful to approximately 25 feet or less in depth</li> <li>Vacuum can cause loss of dissolved gases and volatile organics</li> <li>Pump must be primed and vacuum is often difficult to maintain</li> <li>May cause pH modification</li> </ul>                                    |
| Inertia Pump                    | <ul> <li>Portable, inexpensive, and readily available</li> <li>Rapid method for purging relatively shallow wells</li> </ul>   | <ul> <li>Only useful to approximately 70 feet or less in depth</li> <li>May be time consuming to use</li> <li>Labor intensive</li> <li>WaTerra pump is only effective in 2-inch diameter wells</li> </ul>  |